

**The Infectious Diseases Laboratory and
The Emerging Diseases
Research Group For Companion Animals
at the
University of Georgia
College of Veterinary Medicine**



PDD Sample Submission Protocol For Veterinarians

This protocol offers a complete listing of the samples necessary for full documentation and testing parameters of our PDD research. Alternatives to the protocol, if optimal sample collection cannot be done, are possible. *Please note that the samples can come from either live birds or necropsy specimens.*

Prior to sample collection, please email the Infectious Diseases Laboratory (IDL) at the University of Georgia College of Veterinary Medicine IDLLAB@uga.edu (*Please enter "Inquiry" [without the quotation marks] in the subject heading. Emails without "Inquiry" are treated as spam and irrevocably discarded*) for shipment of a specimen collection kit for non-formalin preservation of tissues (the tubes in the kit are highlighted in bold below; formalin is NOT provided) and to discuss alternative sample collection.

Collection of specimens in the IDL collection kit is highly recommended; however, for circumstances that preclude waiting for the kit, and alternative is to immediately freeze freshly collected samples that would have otherwise been placed in the special fixative. As a rule of thumb, the colder, the better!

Please include name, address, and phone# in the body of the email. Alternatively, this information can be submitted online via the Avian Health Network: <http://stopppdd.org/contactahnform.htm> or request@stopppdd.org (you will receive a reply that requests verification of the authenticity of the sender. Another method to reduce spam!

Samples should include:

1. Two whole blood samples:
Whole blood in EDTA and whole blood in EDTA placed in **fixative solution**;
2. Two fecal samples:
cloacal swabs or fecal samples (feces are preferred): frozen and in **fixative solution**;
3. Three samples of each tissue: *
 - a. fixed in **special fixative**;
 - b. fixed in formalin;
 - c. And frozen.

**Please note: Biopsy or necropsy tissues are needed to confirm PDD (an alternative is sending a previous histopathology slide or pathology report that confirmed PDD).*

Minimum tissues are:

for antemortem biopsies: crop;
for postmortem tissue collection: brain, proventriculus, ventriculus, and crop.

4. Samples should be distributed as follows:

a. Whole blood:

collect in tube containing EDTA anticoagulant (NOT heparin); transfer ½ of the blood from this tube (maximum of 500 ul) into the **tube labeled "blood"**

b. Feces/cloacal swab:

place feces (approximately the size of a pencil eraser) in the **tube labeled "feces"**; freeze a similar fecal aliquot
-for cloacal swabs, place the swab in the solution in the **tube labeled "feces"** and snap off excess handle, place stopper in the tube; freeze a second swab.

c. Tissues:

Divide each tissue into quarters. Place 1/4 of each GI tissue (each should be a minimum size of 1/2 of a pencil eraser) in the **tube labeled "GI tissues"**; place 1/4 (minimum ½ pencil eraser) of cerebrum, cerebellum, and brainstem into the **tube labeled "brain"**; the remaining 50% of the GI and brain should be proportioned as follows: one set (25%) fixed in formalin and a second set (25%) frozen. Any ancillary tissues should be fixed in formalin.

Keep samples that were placed in the non-formalin fixative (**collection kit tubes**) at room temperature overnight. Please read the important notes below prior to shipping samples.

Send all samples the next day **on ice** by overnight mail to:

PDD Study
ATTN: Dr. C.R. Gregory/Dr. B.W. Ritchie
Infectious Diseases Laboratory
University of Georgia College of Veterinary Medicine
501 D.W. Brooks Drive
Athens, GA 30602

Important Notes:

Samples must be placed in the fixatives (**sample collection kit tubes** and formalin) IMMEDIATELY after collection. The ratio of samples to the volume of fixative in the **sample collection kit tubes** is critical-specimens greater than ½ cm³ will not fix properly.

DO NOT FREEZE TISSUES that are to be placed in fixatives and do not freeze them *after* fixation. Samples collected with the **sample collection kit** may be refrigerated over the weekend if necessary. Formalin-fixed tissues should remain at room temperature but may be shipped in the container containing the coolant (ice or ice packs). Please place samples in individual Ziploc bags to prevent leakage and contamination by water.

Samples to be frozen should be IMMEDIATELY frozen after collection.

Samples may be shipped to IDL for delivery Tuesday through Friday; DO NOT send samples that will arrive at the UGA College of Veterinary Medicine on Saturday or Sunday. Please contact IDL prior to shipment. Contact information is on page #1.

Please include signalment and history (either outside of the ice/ice pack container or in a sealed Ziploc bag if it to be placed in the container) along with samples.

Thank you for your support of our research!

Emerging Diseases Research Group for Companion Animals
& the
Infectious Diseases Laboratory
University of Georgia College of Veterinary Medicine